MTS™ Piperacillin-tazobactam 0.064/4-1024/4

Technical Sheet

INDICATIONS FOR USE/INTENDED USE

The MTS™ (MIC Test Strip) Piperacillin-tazobactam TZP 0.064/4-1024/4 is a quantitative method intended for the *in vitro* determination of antimicrobial susceptibility of bacteria. MTS™ consists of specialized paper impregnated with a pre-defined concentration gradient of an antimicrobial agent, which is used to determine the minimum inhibitory concentration (MIC) in µg/mL of antimicrobial agents against bacteria as tested on agar media using overnight incubation and manual reading procedures.

Piperacillin/tazobactam has been shown to be active both clinically and *in vitro* against fastidious aerobic and anaerobic bacteria listed below according to the EMA or FDA label for this antimicrobial agent.

MTS™ TZP 0.064/4-1024/4 generates a stable concentration gradient for piperacillin (0.064-1024 µg/mL) in the presence of a fixed concentration of tazobactam (4 µg/mL). It can be used to determine the MIC of Piperacillin-tazobactam against the following microorganisms:

Haemophilus spp.
Anaerobes

DIRECTIONS FOR USE

Storage

Unopened foil packages: On receipt, store MTS™ TZP 0.064/4-1024/4 at –20°C to +8°C until the given expiry date.

Opened foil packages: Leftover MTS™ from an opened foil package (100 strip pack only) must be stored at 2-8°C in the airtight tube, containing desiccant, provided in the pack for no more than 7 days. Do not store near sources of heat and do not expose to excessive temperature variations.

Handling

Before using MTS™ from an unopened package, visually inspect to ensure the package is intact. Do not use the strips if the package has been damaged. When removed from the refrigerator or freezer, allow the package or storage container to reach room temperature for about 30 minutes. Moisture condensing on the outer surface must evaporate completely before opening the package.

Materials Required but Not Provided:

- Agar plate medium (validated by the media manufacturer for use with antimicrobial susceptibility testing, 90 or 150 mm plates)
- Suspension medium
- McFarland Turbidity standard

  (see the guide below for specific instructions)
- Sterile loops, swabs (not too tightly spun), test tubes, pipettes and scissors
- Forceps
- Incubator (35 ± 2°C)
- Quality control organisms
- Additional technical information from **www.liofilchem.com**

Inoculm Preparation

Suspend well-isolated colonies from an overnight agar plate into the suspension medium to achieve the turbidity of the recommended McFarland standard. If the inoculum concentration is correct, a confluent lawn of growth will be obtained after incubation. If insufficient growth occurs, the testing should be repeated. In order to verify that your procedure gives the correct inoculum density in terms of CFU/mL performing regular colony counts is recommended. An acceptable inoculum should give approximately 1-2 x 10^8 CFU/mL.

Inoculation

Dip a sterile swab in the broth culture or in a diluted form thereof and squeeze it on the wall of the test tube to eliminate excess liquid. Streak the swab over the entire sterile agar surface. Repeat this procedure by streaking 2 more times, rotating the plate approximately 60 degrees each time to ensure an even distribution of inoculum. Allow excess moisture to be absorbed so that the surface is completely dry before applying MTS™.

Application

Apply the strip to the agar surface with the scale facing upwards and code of the strip to the outside of the plate, pressing it with a sterile forceps on the surface of the agar and ensure that whole length of the antibiotic gradient is in complete contact with the agar surface. Once applied, do not move the strip.

Incubation

Incubate the agar plates in an inverted position at the appropriate temperature, atmosphere and time.

Application Guide for MTS™ TZP 0.064/4-1024/4		
Organism	*Haemophilus influenzae* and *Haemophilus parainfluenzae*	Anaerobes
Medium	Haemophilus Test Medium (CLSI) or Mueller Hinton Fastidious Agar (EUCAST)	Brucella Blood Agar with Hemin and Vitamin K1
Inoculum	Suspension in broth to 0.5 McFarland	Suspension in broth to 1 McFarland
Incubation	Agar plates in inverted position at 35 ± 2°C in 5% CO_2 in air for 16-20 hours	Agar plates in inverted position at 36 ± 1°C for 24-72 hours in anaerobic atmosphere

Reading the MIC

After the required incubation period, and only when an even lawn of growth is distinctly visible, read the MIC value where the relevant inhibition ellipse intersects the strip. Do not read the plate if the culture appears mixed or if the lawn of growth is too light or too heavy.

For bactericidal drugs like piperacillin-tazobactam, read the MIC endpoint at complete inhibition of growth. Haze and macrocolonies or microcolonies within 3 mm from the strip should be read as growth.

Growth along the entire gradient i.e. no inhibition ellipse indicates that the value is greater than or equal to (≥) the highest value on the scale. An inhibition ellipse that intersects below the lower end of the scale is read as less than (<) the lowest value. Intersection between two scale segments should be rounded up to the higher value. An MIC of 0.125 µg/mL is considered the same as 0.12 µg/mL for reporting purposes.

Results Interpretation
To categorize the result, typically as susceptible, intermediate or resistant, refer to current MIC breakpoints published by the CLSI, EUCAST and/or your national reference group (MIC interpretative criteria for defining categories are shown below). Always round up MTS™ half dilution values to the next upper two-fold value before categorization. For example a *H. influenzae* Piperacillin-tazobactam MIC of 0.75 µg/mL is reported as 1 µg/mL (see reading guide section for example pictures).

Eliminating Used Material
After use, MTS™ and the material that comes into contact with the sample must be decontaminated and disposed of in accordance with current laboratory techniques for the decontamination and disposal of potentially infected material.

CLSI Interpretation			
Organism	MIC Criteria (µg/mL)		
	S ≤	I	R ≥
Haemophilus spp.	1	-	2
Anaerobes	16	32-64	128

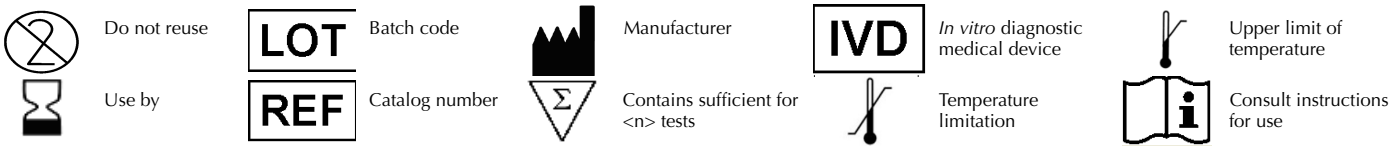
EUCAST Interpretation		
Organism	MIC Criteria (µg/mL)	
	S ≤	R >
H. influenzae	0.25	0.25
Gram-positive anaerobes except *C. difficile*	8	16
Gram-negative anaerobes	8	16

QUALITY CONTROL
CLSI quality control strains are used according to the method as outlined under DIRECTIONS FOR USE.

MIC QC Ranges (µg/mL)	
H. influenzae ATCC® 49247	0.06-0.5
B. fragilis ATCC® 25285	0.12-0.5
B. thetaiotaomicron ATCC® 29741	4-16
E. lenta ATCC® 43055	4-16

REFERENCES

1. CLSI. Performance Standards for Antimicrobial Susceptibility Testing. 30th ed. CLSI supplement M100. Wayne, PA: Clinical and Laboratory Standards Institute; 2020.
2. The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 10.0, 2020. http://www.eucast.org.
3. The European Committee on Antimicrobial Susceptibility Testing. Routine and extended internal quality control for MIC determination and disk diffusion as recommended by EUCAST. Version 10.0, 2020. http://www.eucast.org.
4. CLSI. Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. 11th ed. CLSI standard M07. Wayne, PA: Clinical and Laboratory Standards Institute; 2018.
5. CLSI. Methods for Dilution Antimicrobial Susceptibility Testing of Anaerobic Bacteria. 9th ed. CLSI standard M11. Wayne, PA: Clinical and Laboratory Standards Institute; 2018.
6. ISO 20776-1:2006. Clinical laboratory testing and in vitro diagnostic test systems. Susceptibility testing of infection agents and evaluation of performance of antimicrobial susceptibility test devices—part 1, reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases. ISO, Geneva, Switzerland.

TABLE OF SYMBOLS


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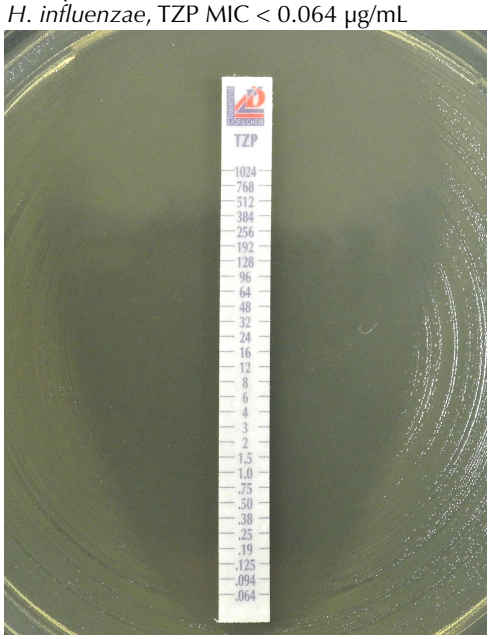
MTS™ Piperacillin-tazobactam Reading Guide

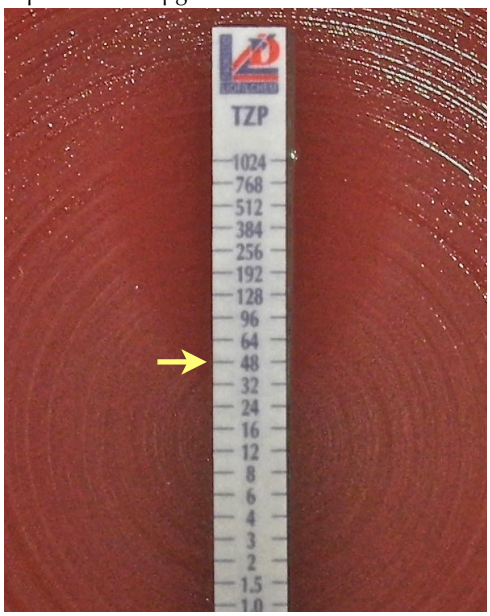
Note: Interpret the MIC as 100% inhibition

Example 1:
H. influenzae, TZP MIC < 0.064 µg/mL



Example 2:
B. fragilis, TZP MIC = 1.5 µg/mL,
reported as 2 µg/mL

Example 3:
B. thetaiotaomicron, TZP MIC = 48 µg/mL,
reported as 64 µg/mL



Example 4:
B. thetaiotaomicron, TZP MIC = 16 µg/mL

Description	µg/mL	Code	Packaging	Ref.
MTS™ Piperacillin-tazobactam	0.064/4 - 1024/4	TZP	10 30 100	921131 92113 921130

MTS™ (MIC Test Strip)
International Patent

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