

MTS[™] Cefiderocol 0.016-256

Technical Sheet

INDICATIONS FOR USE/INTENDED USE

Cefiderocol MTS™ (MIC Test Strip), FDC 0.016-256 µg/mL, is a quantitative method for the *in-vitro* susceptibility testing of *Pseudomonas* aeruginosa to Cefiderocol.

MTS[™] consists of special porous paper impregnated with a pre-defined concentration gradient of an antimicrobial agent, used to determine the minimum inhibitory concentration (MIC) in µg/mL of antimicrobial agents against bacteria as tested on agar media using overnight incubation and manual reading procedures.

Cefiderocol is a siderophore cephalosporin with activity against aerobic Gram-negative organisms. It binds to free iron and is actively transported into bacterial cells through the outer membrane. This Trojan horse strategy allows cefiderocol to enter the space in-between the bacterial cell walls and disrupt cell wall synthesis. In addition, cefiderocol is stable against nearly all beta-lactamases, including both the serine and metallo-carbapenemases.

DIRECTIONS FOR USE

Storage

<u>Unopened foil packages</u>: On receipt, store MTS[™] FDC 0.016-256 at −20°C until the given expiry date.

<u>Opened foil packages</u>: Leftover MTSTM from an opened foil package (100 strip pack only) must be stored at 2-8°C in the airtight tube, containing desiccant, provided in the pack for no more than 7 days. Do not store near sources of heat and do not expose to excessive temperature variations.

Handling

Before using MTS[™] from an unopened package, visually inspect to ensure the package is intact. Do not use the strips if the package has been damaged. When removed from the refrigerator or freezer, allow the package or storage container to reach room temperature for about 30 minutes. Moisture condensing on the outer surface must evaporate completely before opening the package.

Materials Required but Not Provided:

- Agar plate medium (validated by the media manufacturer for use with antimicrobial susceptibility testing, 90 or 150 mm plates)
- Suspension medium
- McFarland Turbidity standard

(see the guide below for specific instructions)

- Sterile loops, swabs (not too tightly spun), test tubes, pipettes and scissors
- Forceps
- Incubator $(35 \pm 2^{\circ}C)$
- Quality control organisms
- Additional technical information from www.liofilchem.com

Inoculm Preparation

Suspend well-isolated colonies from an overnight agar plate into the suspension medium to achieve the turbidity of the recommended McFarland standard. If the inoculum concentration is correct, a confluent lawn of growth will be obtained after incubation. If insufficient growth occurs, the testing should be repeated. In order to verify that your procedure gives the correct inoculum density in terms of CFU/mL performing regular colony counts is recommended. An acceptable inoculum should give approximately 1-2 x 10⁸ CFU/mL.

Inoculation

Dip a sterile swab in the broth culture or in a diluted form thereof and squeeze it on the wall of the test tube to eliminate excess liquid. Streak the swab over the entire sterile agar surface. Repeat this procedure by streaking 2 more times, rotating the plate approximately 60 degrees each time to ensure an even distribution of inoculum. Allow excess moisture to be absorbed so that the surface is completely dry before applying MTSTM.

Application

Apply the strip to the agar surface with the scale facing upwards and code of the strip to the outside of the plate, pressing it with a sterile forceps on the surface of the agar and ensure that whole length of the antibiotic gradient is in complete contact with the agar surface. Once applied, do not move the strip.

Incubation

Incubate the agar plates in an inverted position at the appropriate temperature, atmosphere and time.

Application Guide for MTS™ FDC 0.016-256				
Organism	Pseudomonas aeruginosa			
Medium	Mueller Hinton Agar ¹			
Inoculum	Suspension in saline (0.85% NaCl) to 0.5 McFarland (1 if mucoid)			
Incubation	Agar plates in inverted position at $35 \pm 2^{\circ}$ C for 16-20 hours in ambient atmosphere			

Cefiderocol requires low iron levels for optimal activity. Standard cation-adjusted Mueller-Hinton agar (MHA) is not controlled for iron concentration and iron concentrations may vary depending on the manufacturer. MTSTM Cefiderocol FDC 0.016-256 has been tested and validated with MHA from Liofilchem and BD.

Reading the MIC

After the required incubation period, and only when an even lawn of growth is distinctly visible, read the MIC value where the relevant inhibition ellipse intersects the strip. Do not read the plate if the culture appears mixed or if the lawn of growth is too light or too heavy. For bactericidal drugs like cefiderocol, read the MIC endpoint at complete inhibition of growth. Haze and macrocolonies or microcolonies within 3 mm from the strip should be read as growth.

Growth along the entire gradient i.e. no inhibition ellipse indicates that the value is greater than or equal to \geq the highest value on the scale. An inhibition ellipse that intersects below the lower end of the scale is read as less than (<) the lowest value. Intersection between two scale segments should be rounded up to the higher value. An MIC of 0.125 µg/mL is considered the same as 0.12 µg/mL for reporting purposes.

Results Interpretation

To categorize the result, typically as susceptible, intermediate or resistant, refer to current MIC breakpoints published by the CLSI, EUCAST and/or your national reference group (MIC interpretative criteria for defining categories are shown below). Always round up MTSTM half dilution values to the next upper two-fold value before categorization. For example a *P. aeruginosa* Cefiderocol MIC of 0.75 µg/mL is reported as 1 µg/mL (see reading guide section for example pictures).

Eliminating Used Material

After use, MTSTM and the material that comes into contact with the sample must be decontaminated and disposed of in accordance with current laboratory techniques for the decontamination and disposal of potentially infected material.

QUALITY CONTROL

Quality control strains recommended by CLSI and EUCAST are used according to the method as outlined under DIRECTIONS FOR USE.

	MIC Breakpoints (µg/mL)						
Organism	CLSI			EUCAST		MIC QC Ranges (µg/mL)	
	S ≤		R ≥	S ≤	R >		
Pseudomonas aeruginosa	4	8	16	2	2	P. aeruginosa ATCC [®] 27853 0.06-0.5	

PERFORMANCE CHARACTERISTICS

The performance of MTSTM FDC 0.016-256 has been established by comparison to the broth microdilution (BMD) reference method following CLSI M07 and ISO 20776-1 standards. Performance was evaluated using the following indices: essential agreement (EA), category agreement (CA), minor error (mE), major error (ME), and very major error (VME).

Organism	Ν	% Essential Agreement	% Category Agreement		
Organism		% Essential Agreement	(CLSI breakpoints)	(EUCAST breakpoints)	
Pseudomonas aeruginosa	300	90.3	97.3	99.3	

Essential agreement (EA) was defined as agreement between MTS[™] and BMD methods ± 1 doubling dilution.

The following errors resulted when MTS™ FDC MICs were 1 doubling dilution apart from the BMD MICs:

- Applying CLSI breakpoints, 8 minor errors (mEs) were observed. No very major errors (VMEs) nor major errors (MEs) were found;
- Applying EUCAST breakpoints, 1 VME and 1 ME were found. No mEs were found.

REFERENCES

- 1. CLSI. Performance Standards for Antimicrobial Susceptibility Testing. 31st ed. CLSI supplement M100. Wayne, PA: Clinical and Laboratory Standards Institute; 2021.
- 2. The European Committee on Antimicrobial Susceptibility Testing. Breakpoint tables for interpretation of MICs and zone diameters. Version 11.0, 2021. http://www.eucast.org.
- 3. The European Committee on Antimicrobial Susceptibility Testing. Routine and extended internal quality control for MIC determination and disk diffusion as recommended by EUCAST. Version 11.0, 2021. http://www.eucast.org.
- 4. CLSI. Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. 11th ed. CLSI standard M07. Wayne, PA: Clinical and Laboratory Standards Institute; 2018.
- 5. ISO 20776-1:2006. Clinical laboratory testing and in vitro diagnostic test systems. Susceptibility testing of infection agents and evaluation of performance of antimicrobial susceptibility test devices—part 1, reference method for testing the in vitro activity of antimicrobial agents against rapidly growing aerobic bacteria involved in infectious diseases. ISO, Geneva, Switzerland.

TABLE OF SYMBOLS



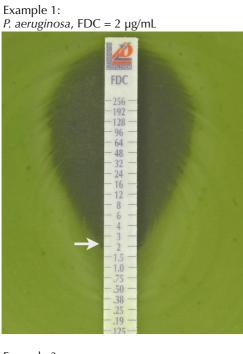
CLSI is a trademark belonging to Clinical Laboratory and Standards Institute, Inc.

The ATCC trademark and trade name and any and all ATCC catalog numbers are trademarks of the American Type Culture Collection. This document has been produced in part under ECDC service contracts and made available at no cost by EUCAST at no cost to the user and can be accessed on the EUCAST website: www.eucast.org. EUCAST recommendations are frequently updated and the latest versions are available at www.eucast.org.

Any other name or trademark is the property of its respective owner.

MTS[™] Cefiderocol Reading Guide

Note: Interpret the MIC as 100% inhibition



Example 3: P. aeruginosa, FDC MIC = 0.25 µg/mL



Example 2: *P. aeruginosa*, FDC MIC = $0.75 \mu g/mL$, reported as $1 \mu g/mL$



Example 4: P. aeruginosa, FDC MIC = 0.25 µg/mL



Description	µg/mL	Code	Packaging	Ref.
MTS™ Cefiderocol	0.016 - 256	FDC	10 30 100	920671 92067 920670

MTS™ (MIC Test Strip) International Patent

This document is available from liofilchem.com/MTS

Liofilchem®, the Liofilchem company logo and MTS logo are registered trademarks of LIOFILCHEM s.r.l.



LIOFILCHEM[®] s.r.l.

Via Scozia, 64026 Roseto degli Abruzzi (TE) Italy Tel. +39 0858930745 Fax +39 0858930330

www.liofilchem.com

