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Liofilchem® MTS™

Rx only

IVD

SUMMARY AND EXPLANATION OF THE TEST

The Liofilchem® MTS™ (MIC Test Strip) are gradient tests used to determine the minimum inhibitory concentration (MIC) of select bacteria to indicate appropriate patient treatment and for identifying resistance patterns. The MIC is the minimum inhibitory concentration of an antibiotic that will inhibit the growth of bacteria under standardized *in vitro* conditions. Broth and agar dilution MIC procedures based on two-fold serial dilutions of antibiotics are the reference methodologies; expected reproducibility of which is within ± 1 two-fold dilution (1).

PRINCIPLE OF THE METHOD

MTS™ are made of special high quality paper impregnated with a predefined concentration gradient of antibiotic, across 15 two-fold dilutions like those of a conventional MIC method. When the MTS™ is applied onto an inoculated agar surface, the preformed exponential gradient of antimicrobial agent diffuses into the agar for over an hour. After incubation, a symmetrical inhibition ellipse centered along the strip is formed. The MIC is read directly from the scale in terms of µg/mL at the point where the edge of the inhibition ellipse intersects the strip MTS™.

REAGENTS

MTS™ is supplied in 3 different packaging options (no additional reagents are included):

- The 10-test box contains 10 strips individually packed in desiccant envelopes.
- The 30-test box contains 30 strips individually packed in desiccant envelopes.
- The 100-test box contains 100 strips in a canister with a desiccant built into the lid.

This instruction sheet is available from www.liofilchem.com/MTS/US

DIRECTIONS FOR USE

Storage

Unopened foil packages and canisters: On receipt, store MTS™ at -20°C to $+8^{\circ}\text{C}$ until the given expiry date. Some MTS™ (e.g. carbapenems) should be stored frozen at -20°C . Check the drug-specific MTS™ supplement for the specific storage temperature.

Opened canisters: MTS™ in canister can be used for up to 2 months from first opening (record the date on which the canister was open) and must be stored at the label storage temperature. Before using the remaining strips, check the expiry date indicated on the packaging. Do not store near sources of heat and do not expose to excessive temperature variations.

Protect MTS™ from moisture, heat and direct exposure to strong light at all times.

Handling

Before using the MTS™ from an unopened package, visually inspect to ensure the package is intact. Do not use the strips if the package has been damaged. When removed from the refrigerator/freezer, allow the package or storage container to reach room temperature for about 30 minutes. Moisture condensing on the outer surface must evaporate completely before opening the package. Use forceps or a similar device to pick up a strip.

When using MTS™ from a canister, replace the lid immediately after use and store as outlined under STORAGE.

Precautions

The MTS™ is not classified as being hazardous according to current regulations. The MTS™ is a disposable product. The MTS™ is only for diagnostic *in vitro* use and is intended for professional use. They must be used in the laboratory by properly trained operators using approved aseptic and safety methods for pathogenic agents.

Per the FDA-Recognized Susceptibility Test Interpretive Criteria website, the safety and efficacy of antimicrobial drugs, for which antimicrobial susceptibility is tested by this AST device, may or may not have been established in adequate and well-controlled clinical trials for treating clinical infections due to microorganisms outside of those found in the indications and usage in the drug label. The clinical significance of susceptibility information in those instances is unknown. The approved labeling for specific antimicrobial drugs provides the uses for which the antimicrobial drug is approved.

Materials Required but Not Provided:

- Agar plate medium (validated by the media manufacturer for use with antimicrobial susceptibility testing, 90 or 150 mm plates)
 - Suspension medium
 - McFarland turbidity standard
 - Sterile loops, swabs (not too tightly spun), test tubes, pipettes and scissors
 - Forceps
 - Incubator ($35 \pm 2^{\circ}\text{C}$)
 - Quality control organisms
 - Additional technical information from www.liofilchem.com
- (The medium to be used as well as the inoculum suspension will depend on the organism under investigation, see the MTS™ Supplement for more information)

Inoculum Preparation

Suspend well-isolated colonies from an overnight agar plate into the suspension medium to achieve the turbidity of the recommended McFarland standard. If the inoculum concentration is correct, a confluent lawn of growth will be obtained after incubation. If insufficient growth occurs, the testing should be repeated.

McFarland turbidity standards do not guarantee the correct number of viable cells in the suspension. In order to verify that your procedure gives the correct inoculum density in terms of CFU/mL performing regular colony counts is recommended. An acceptable inoculum should give approximately $1-2 \times 10^8$ CFU/mL.

Inoculation

Dip a sterile swab in the broth culture or in a diluted form thereof and squeeze it on the wall of the test tube to eliminate excess liquid. Streak the swab over the entire sterile agar surface. Repeat this procedure by streaking 2 more times, rotating the plate approximately 60 degrees each time to ensure an even distribution of inoculum. Allow excess moisture to be absorbed so that the surface is completely dry before applying MTS™.

Use well-defined, high quality media for AST that supports good growth. The brand chosen should have good batch-to-batch reproducibility to ensure that accurate and reliable MIC values are obtained.

The agar medium should have a depth of 4.0 ± 0.5 mm, a pH of 7.3 ± 0.1 and all other quality specifications should be fulfilled. Refer to the media manufacturer's instructions for more information.

Application

Apply the strip to the agar surface with the scale facing upwards and code of the strip to the outside of the plate, pressing it with sterile forceps on the surface of the agar and ensure that whole length of the antibiotic gradient is in complete contact with the agar surface. Once applied, do not move the strip.

Incubation

Incubate the agar plates in an inverted position at the appropriate temperature, atmosphere and time. Refer to the drug-specific MTS™ Supplement for specific incubation instructions.

Reading the MIC

After the required incubation period, and only when an even lawn of growth is distinctly visible, read the MIC value where the relevant inhibition ellipse intersects the strip. Do not read the plate if the culture appears mixed or if the lawn of growth is too light or too heavy.

NOTES:

- Antimicrobial drugs can be either “-static” (e.g. bacteriostatic, fungistatic) or “-cidal” in their interactions with target organisms and this needs to be considered for determining correctly the MIC endpoint. For bactericidal drugs, e.g. beta-lactams, read the MIC at the point of complete inhibition of all growth. Haze and macrocolonies or microcolonies within 3 mm from the strip should be read as growth. For bacteriostatic drugs, e.g. trimethoprim-sulfamethoxazole, in case of trailing endpoints, read at 80% inhibition, i.e. the first point of significant inhibition as judged by the naked eye. Consult MTS30 (cidal-static technical sheet) for more information.
- Growth along the entire gradient i.e. no inhibition ellipse indicates that the value is greater than or equal to (\geq) the highest value on the scale. An inhibition ellipse that intersects below the lower end of the scale is read as less than ($<$) the lowest value. Intersection between two scale segments should be rounded up to the higher value. An MIC of 0.125 µg/mL is considered the same as 0.12 µg/mL for reporting purposes. See the appropriate MTS™ technical sheets for example specific drug-organism photographs. Also consult the MTS™ Photographic Guide.
- Excessively wet plates prior to inoculation, insufficient drying before applying strips and/or unevenly streaked surfaces may give non confluent growth or jagged ellipse edges. Repeat the test if MIC endpoints are difficult to read. In the case of uneven MIC intersections, read the higher value. Repeat the test if the discrepancy is >1 dilution.
- Occasionally, certain antimicrobial agent/microorganism combinations may give unusual results. In these cases, judgment of the MIC endpoint may be difficult for the inexperienced personnel. However, individuals can be trained through regular use of quality control strains, MTS™ reading guides and comparison with experienced personnel to correctly assess MIC endpoints.

Result Interpretation

To categorize the result according to the interpretive criteria, refer to the appropriate MTS™ product supplement for the specific antimicrobial agent interpretive criteria. Since MTS™ generates MIC values which fall between two-fold dilutions for interpretation, an MTS™ MIC value which falls between standard two-fold dilutions must be rounded up to the next standard upper two-fold value before categorization. For example a *S. aureus* vancomycin MIC of 1.5 µg/mL is reported as 2 µg/mL.

NOTES:

- As with all AST data, MTS™ results are *in vitro* values only and may provide an indication of the organism's potential *in vivo* susceptibility. The use of results to guide therapy selection must be the sole decision and responsibility of the attending physician. Their judgement should be based on the medical history and knowledge of the patient, pharmacokinetics/pharmacodynamics of the antimicrobial agent, and clinical experience in treating infections caused by the particular microbial pathogen. The drug, dose and dosing regimen must also be considered.
- For details of specific interpretive limitations and/or limitations on the clinical use of an antimicrobial agent in various therapeutic situations, please refer to the tables and footnotes of MIC interpretive standards in the latest CLSI documents.

Eliminating Used Material

After use, MTS™ and the material that comes into contact with the sample must be decontaminated and disposed of in accordance with current laboratory techniques for the decontamination and disposal of potentially infected material.

QUALITY CONTROL

To check the performance of the MTS™ result, test the quality control strain(s) as shown in the appropriate MTS™ product supplement. Patient isolate results are considered satisfactory if the quality control result(s) fall within the expected range(s). Patient isolate results should not be reported if the quality control results are outside of this stated QC range. MIC results for a QC strain that fall a half dilution below the lower QC limit should be rounded up to the next upper two-fold value which would establish QC compliance. MIC results that are a half dilution above the upper limit would be rounded up to the next upper two-fold value which would result in non-QC compliance.

LIMITATIONS

Refer to the drug-specific MTS™ Supplement.

EXPECTED VALUES

Expected results for susceptibility tests will vary based on location and institution. Organism resistance patterns will be directly related to the population of organisms at each site.








PERFORMANCE CHARACTERISTICS

Refer to the drug-specific MTS™ Supplement.

REFERENCES

1. CLSI. *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically*. 11th ed. CLSI standard M07. Wayne, PA: Clinical and Laboratory Standards Institute; 2018.

GLOSSARY OF TERMS

 Do not reuse	LOT Batch code	 Manufacturer	IVD <i>In vitro</i> diagnostic medical device	 Upper limit of temperature
 Use by	REF Catalog number	 Contains sufficient for <n> tests	 Temperature limitation	 Consult instructions for use



Drug Specific Supplement for MTS™ Omadacycline

Rx only

IVD

Indications for Use/Intended Use

The MTS™ (MIC Test Strip) Omadacycline 0.002-32 µg/mL is a quantitative method intended for the *in vitro* determination of antimicrobial susceptibility of bacteria. MTS™ consists of specialized paper impregnated with a pre-defined concentration gradient of an antimicrobial agent, which is used to determine the minimum inhibitory concentration (MIC) in µg/mL of antimicrobial agents against bacteria as tested on agar media using overnight incubation and manual reading procedures. The MTS™ Omadacycline at concentrations of 0.002 – 32 µg/mL should be interpreted at 16-20 hours (non-fastidious organisms) and 20-24 hours (fastidious organisms) of incubation.

Omadacycline has been shown to be active both clinically and in vitro against these bacterial species according to the FDA drug approved label:

Gram-positive bacteria

Staphylococcus aureus
Staphylococcus lugdunensis
Enterococcus faecalis
Streptococcus pneumoniae
Streptococcus pyogenes
Streptococcus anginosus group (includes *S. anginosus* and *S. constellatus*)

Gram-negative bacteria

Enterobacter cloacae
Klebsiella pneumoniae
Haemophilus influenzae
Haemophilus parainfluenzae

Omadacycline has been shown to be active in vitro only against the bacterial species listed below according to the FDA drug approved label:

Gram-positive bacteria

Enterococcus faecium (vancomycin-susceptible and -resistant isolates)

Gram-negative bacteria

Escherichia coli
Citrobacter freundii
Citrobacter koseri
Klebsiella (Enterobacter) aerogenes
Klebsiella oxytoca

Specifications

Antibiotic code: OMC

MIC range: 0.002-32 µg/mL

Antibiotic group: Aminomethylcycline

Directions for Use

Follow the MTS™ package insert instructions.

Procedures specific to MTS™ Omadacycline:

Storage	Temperature between –20°C and +8°C		
Organism	<i>Enterobacteriaceae</i> , <i>S. aureus</i> , <i>S. lugdunensis</i> , <i>E. faecalis</i> , <i>E. faecium</i>	<i>H. influenzae</i> , <i>H. parainfluenzae</i>	<i>S. pneumoniae</i> , <i>S. pyogenes</i> , <i>S. anginosus</i> , <i>S. constellatus</i>
Medium	Mueller Hinton Agar	Haemophilus Test Medium	Mueller Hinton + 5% sheep blood Agar
Inoculum	Suspension in saline (0.85% NaCl) to 0.5 McFarland standard	Suspension in saline (0.85% NaCl) to 0.5 McFarland standard (1 if mucoid)	
Incubation	Agar plates in inverted position at 35 ± 2°C for 16-20 hours in ambient atmosphere	Agar plates in inverted position at 35 ± 2°C for 20-24 hours in 5% CO ₂	
Reading	Interpret the MIC as 80% inhibition when trailing is seen	Interpret the MIC as 100% inhibition	

FDA omadacycline interpretive criteria (µg/mL)

Use the following breakpoints to categorize the result according to the interpretive criteria (i.e. susceptible or resistant). An MTS™ MIC value which falls between standard two-fold dilutions must be rounded up to the next standard upper two fold value before categorization. For example a *K. pneumoniae* omadacycline MIC of 0.19 µg/mL is reported as 0.25 µg/mL (see reading guide section for example pictures).

Bacterial Species	Acute Bacterial Skin and Skin Structure Infections (ABSSSI)			Community-Acquired Pneumonia Infections (CABP)		
	Susceptible	Intermediate	Resistant	Susceptible	Intermediate	Resistant
<i>Enterobacteriaceae</i>	≤4	8	≥16	≤4	8	≥16
<i>Enterococcus faecalis</i>	≤0.25	0.5	≥1	-	-	-
<i>Staphylococcus aureus</i>	≤0.5	1	≥2	≤0.25 ¹	0.5 ¹	≥1 ¹
<i>Staphylococcus lugdunensis</i>	≤0.12	0.25	≥0.5	-	-	-

Bacterial Species	Acute Bacterial Skin and Skin Structure Infections (ABSSSI)			Community-Acquired Pneumonia Infections (CABP)		
	Susceptible	Intermediate	Resistant	Susceptible	Intermediate	Resistant
<i>Haemophilus</i> species ²	-	-	-	≤2	4	≥8
<i>Streptococcus pneumoniae</i>	-	-	-	≤0.12	0.25	≥0.5
<i>Streptococcus pyogenes</i>	≤0.12	0.25	≥0.5	-	-	-
<i>Streptococcus anginosus</i> group ³	≤0.12	0.25	≥0.5	-	-	-

¹ Methicillin-susceptible isolates only.

² *Haemophilus* species includes *H. influenzae* and *H. parainfluenzae*.

³ Interpretive criteria for *Streptococcus anginosus* group includes *S. anginosus*, *S. intermedius*, and *S. constellatus*.

US FDA Susceptibility Interpretive Criteria (STIC) Ref:

<https://www.fda.gov/STIC>

Quality Control range (µg/mL) (CLSI M100S Performance Standards for Antimicrobial Susceptibility Testing, 30th Edition)

To check the performance of the MTSTTM Omadacycline, media and procedure, test *E. coli* ATCC 25922 for non-fastidious Gram-negative bacteria, test *E. faecalis* ATCC 29212 and *S. aureus* ATCC 29213 for non-fastidious Gram-positive bacteria, test *S. pneumoniae* ATCC 49619 for fastidious Gram-positive bacteria, and test *H. influenzae* ATCC 49247 for fastidious Gram-negative bacteria according to the method as outlined in the MTSTTM package insert. Results are considered satisfactory if they fall within the following ranges:

Quality Control Strain	Acceptable MIC Range (µg/mL)
<i>Escherichia coli</i> , ATCC [®] 25922	0.25 – 2
<i>Enterococcus faecalis</i> , ATCC [®] 29212	0.06 – 0.5
<i>Staphylococcus aureus</i> , ATCC [®] 29213	0.12 – 1
<i>Streptococcus pneumoniae</i> , ATCC [®] 49619	0.016 - 0.12
<i>Haemophilus influenzae</i> , ATCC [®] 49247	0.5 - 2

Performance Characteristics

Correlation to Reference Method¹

		N	% Essential Agreement	% Category Agreement
Non-Fastidious	<i>Enterobacteriaceae</i>	578	99.7	95.0
	<i>Staphylococcus aureus</i> (ABSSSI)	175	98.9	96.0
	<i>Staphylococcus aureus</i> (CABP)	175	98.9	96.6
	<i>Staphylococcus lugdunensis</i> ²	70	100	87.1
	<i>Enterococcus faecalis</i>	121	98.3	84.3
	<i>Enterococcus faecium</i>	100	100	98.0
	<i>Enterococcus</i> spp.	221	99.1	90.5
Fastidious	<i>Haemophilus influenzae</i>	112	98.2	97.3
	<i>Haemophilus parainfluenzae</i>	30	100	93.3
	<i>Streptococcus pneumoniae</i> ³	181	91.7	96.7
	<i>Streptococcus anginosus</i> group ^{3,4,5}	45	100	77.8
	<i>Streptococcus pyogenes</i> ³	35	100	91.4

¹ For the plate inoculation procedure, one testing site utilized a plate rotator (Retro C80) to assist even distribution of inoculum. There was no difference in performance for the site using the plate rotator as compared to sites using the manual plate inoculation method.

² MTSTTM Omadacycline MIC values tend to be in exact agreement or at least one double dilution higher when testing *S. lugdunensis* compared to the CLSI reference broth microdilution.

³ MTSTTM Omadacycline MIC values tended to be in exact agreement or at least one doubling dilution higher when testing *S. anginosus*, *S. constellatus*, *S. pneumoniae*, and *S. pyogenes* compared to the CLSI reference broth microdilution.

⁴ Includes 27 *S. anginosus* and 18 *S. constellatus*.

⁵ Omadacycline should be tested with *Streptococcus anginosus* and *Streptococcus constellatus* isolates only. The performance of *Streptococcus intermedius* has not been established during the clinical study.

Reproducibility

100% of MTSTTM Omadacycline results for non-fastidious Gram-negative bacteria (4 *E. coli*, 3 *K. pneumoniae* and 3 *E. cloacae* tested in triplicate at 3 sites on 3 days) were within a doubling dilution of reference broth microdilution results. 95.2% of MTSTTM Omadacycline results for non-fastidious Gram-positive bacteria (4 *S. aureus* [2 MSSA, 2 MRSA], 4 *E. faecalis* [2 VSE, 2 VRE] and 2 *S. lugdunensis* tested in triplicate at 3 sites on 3 days) were within a doubling dilution of reference broth microdilution results. 99.3% of MTSTTM Omadacycline results for fastidious Gram-negative and Gram-positive bacteria (2 *H. influenzae*, 1 *H. parainfluenzae*, 3 *S. pneumoniae*, 2 *S. pyogenes*, 2 *S. anginosus*, and 1 *S. constellatus* tested in triplicate at 3 sites on 3 days) were within a doubling dilution of reference broth microdilution results.

Limitations

The ability of MTSTTM to detect non-susceptible isolates with the following drug/bacterial species combinations is unknown because resistant isolates were either not available or an insufficient number was encountered at the time of comparative testing:

Omadacycline: *Citrobacter freundii*, *H. influenzae*, *H. parainfluenzae*, *S. anginosus*, *S. constellatus* and *S. pyogenes*.

The safety and efficacy of omadacycline in treating Acute Bacterial and Skin Structure Infections (ABSSSI) infections due to Gram-negative organisms other than *K. pneumoniae* and *E. cloacae* and Gram-positive organisms other than *S. aureus* (MRSA and MSSA), *S. lugdunensis*, *E. faecalis*, *S. pyogenes*, and *S. anginosus* group may or may not have been established in adequate and well-controlled clinical trials. The clinical significance of susceptibility information in such instances is unknown.

The safety and efficacy of omadacycline in treating Community-Acquired Bacterial Pneumonia (CABP) infections due to Gram-negative organisms other than *K. pneumoniae* and Gram-positive organisms other than *S. aureus* (MSSA only) and *S. pneumoniae* may or may not have been established in adequate and well-controlled clinical trials. The clinical significance of susceptibility information in such instances is unknown.

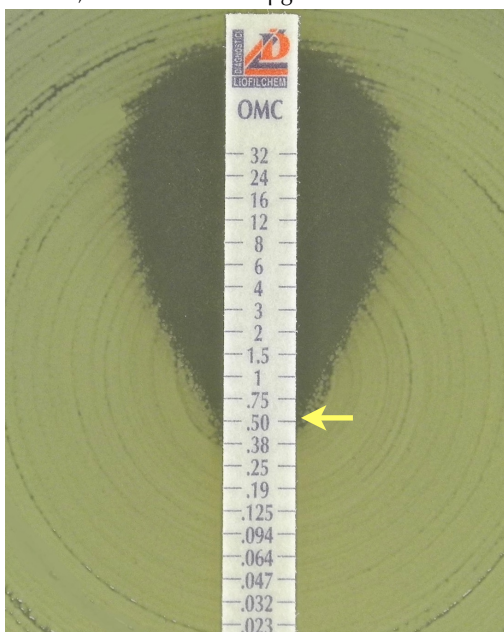
Resistance mechanism characterization was not available for all organisms at the time of comparative testing, and therefore the performance of the MTS™ Omadacycline for non-fastidious and fastidious Gram-negative bacilli and Gram-positive cocci is unknown for the following: Enterobacteriaceae [tet(B)]; *Enterococcus* species [tet(K), tet(L)]; *S. aureus* [tet(L)]; *S. pneumoniae* [tet(K), tet(L), tet(M)]; *S. pyogenes* [tet(K), tet(L), tet(M)]; *S. anginosus* [tet(K), tet(L), tet(M)]; *S. constellatus* [tet(K), tet(L), tet(M)]; *H. influenzae* [tet(B)]; *H. parainfluenzae* [tet(B)].

Omadacycline is not active *in vitro* against *Morganella* spp., *Proteus* spp., and *Providencia* spp.

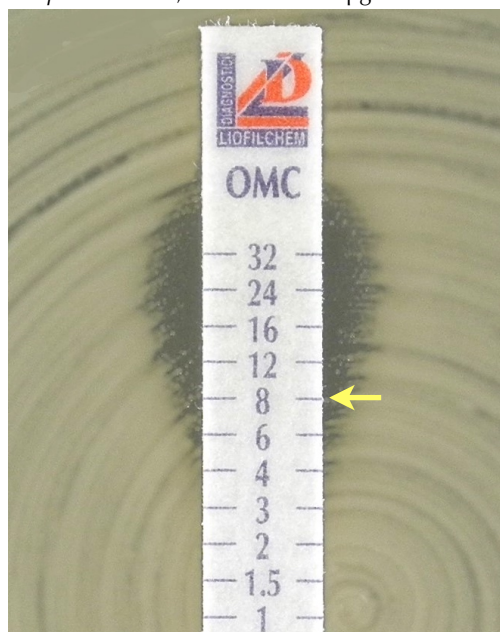
MTS™ Omadacycline Reading Guide

NOTE: Interpret the MIC as 80% inhibition for non-fastidious species

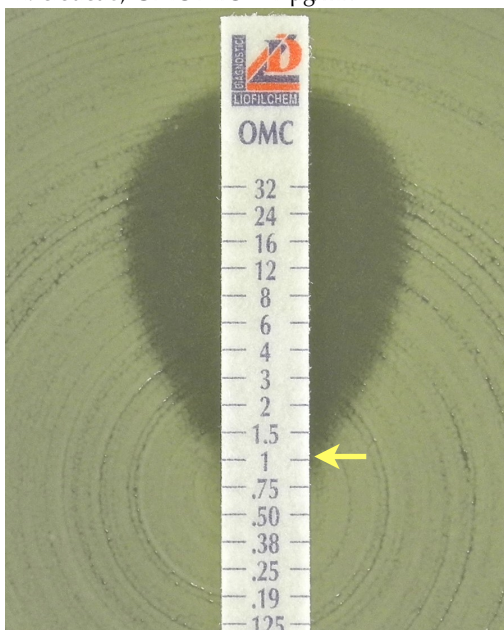
Example 1:
E. coli, OMC MIC = 0.5 µg/mL



Example 2:
K. pneumoniae, OMC MIC = 8 µg/mL



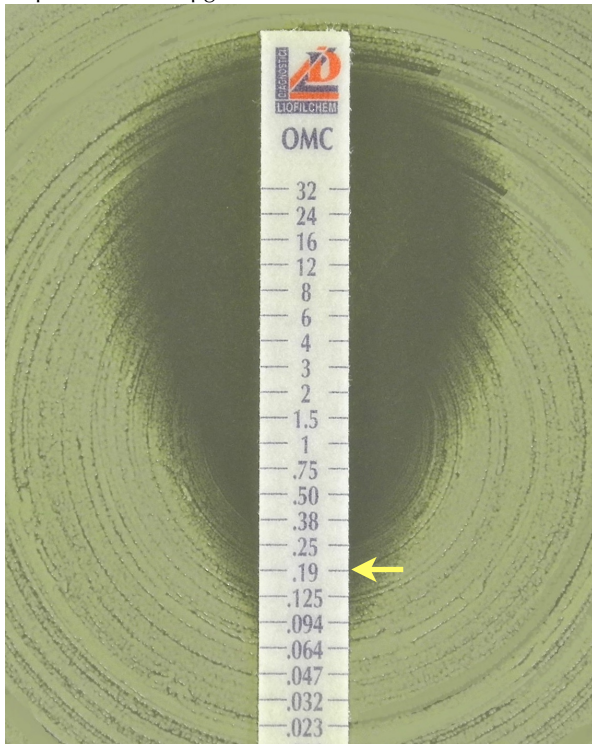
Example 3:
E. cloacae, OMC MIC = 1 µg/mL



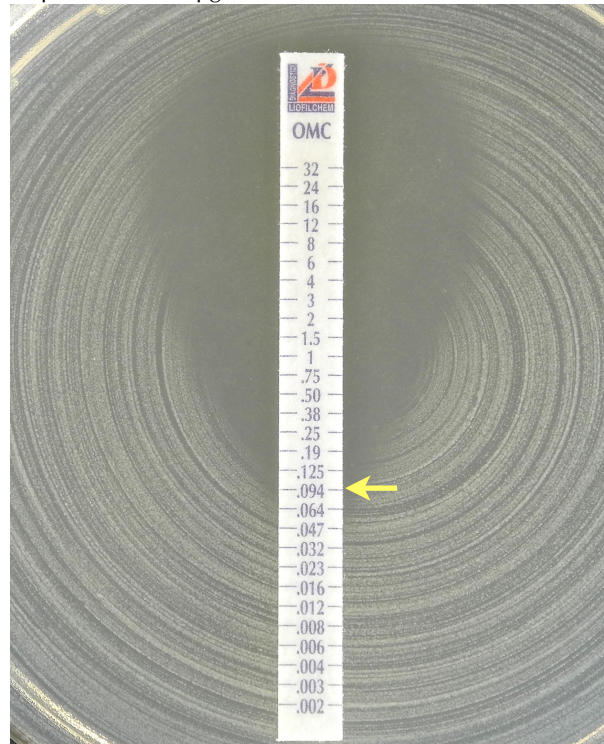
Example 4:
C. freundii, OMC MIC = 1 µg/mL



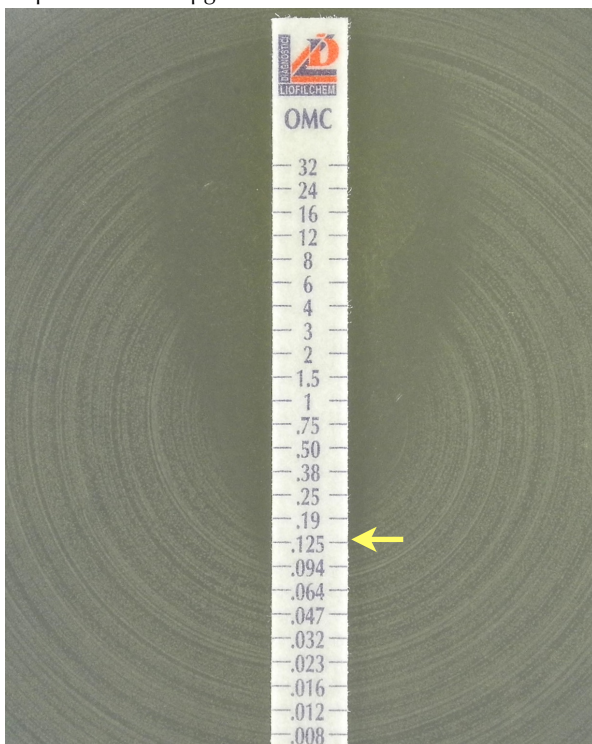
Example 5:
S. aureus, OMC MIC = 0.19 µg/mL,
reported as 0.25 µg/mL



Example 6:
S. lugdunensis, OMC MIC = 0.094 µg/mL,
reported as 0.12 µg/mL



Example 7:
E. faecalis, OMC MIC = 0.125 µg/mL,
reported as 0.12 µg/mL



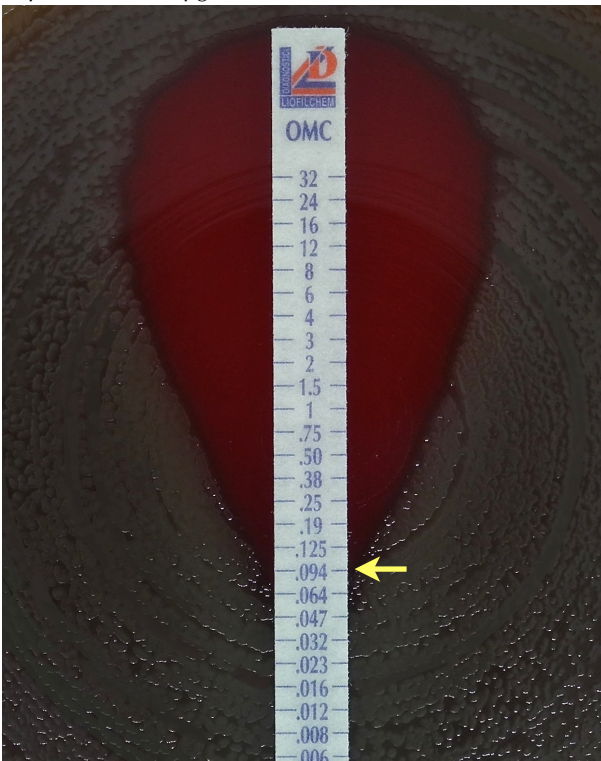
Example 8:
E. faecium, OMC MIC = 0.125 µg/mL,
reported as 0.12 µg/mL



NOTE: Interpret the MIC as 100% inhibition for fastidious species

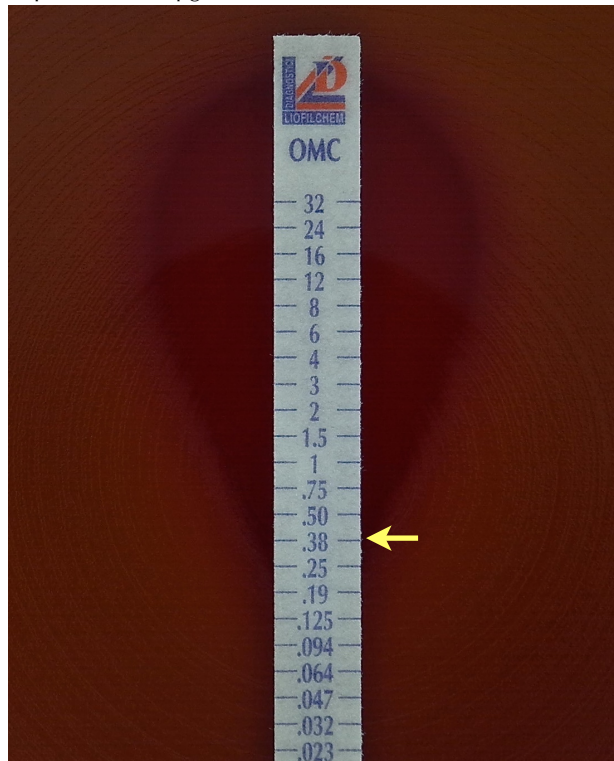
Example 9:

S. pneumoniae, OMC MIC = 0.094 µg/mL,
reported as 0.12 µg/mL



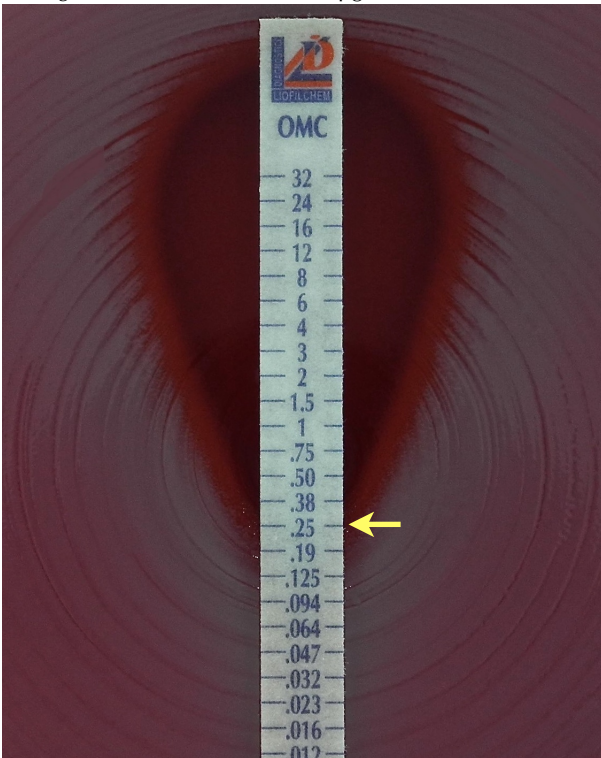
Example 10:

S. pyogenes, OMC MIC = 0.38 µg/mL,
reported as 0.5 µg/mL



Example 11:

S. anginosus, OMC MIC = 0.25 µg/mL



Example 12:

H. influenzae, OMC MIC = 2 µg/mL



PRESENTATION		µg/mL	Code	Packaging	Ref.
MTS™	Omadacycline	0.002 - 32	OMC	10	920711
				30	92071
				100	920710

REVISION HISTORY

Document	Release Date	Change Summary
Package Insert 92071 PI-0 F00518 v5.3-v0 MTS OMC US	2018-12-20	Not applicable (Initial release)
Package Insert 92071 PI-1 F00518 v6.3-v1 MTS OMC US	2020-03-20	Revised: Precautions, Inoculum Preparation Added: Susceptibility testing of fastidious organisms Updated: Interpretive Criteria (Link to FDA-recognized STIC), Quality Control range (Reference to CLSI M100S ED30)
eIFU 92071 IFU-2 MTS OMC US	2022-04-19	Revised: Reagent, Storage and Handling (new Canister packaging), Inoculum Preparation, Inoculation, Reading the MIC, Results Interpretation, Directions for Use (agar plate medium for <i>H. influenzae</i> and <i>H. parainfluenzae</i>) Added: Table of Contents, Revision History

Note: Minor typographical, grammar, and formatting changes are not included in the revision history.

For all inquiries please fill out the form at <https://www.liofilchem.com/contact-us.html>

MTS™ (MIC Test Strip), European Patent

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Liofilchem Inc., US Distributor and Customer Service

465 Waverley Oaks Rd, Waltham, MA
Phone: 781-902-0312



LIOFILCHEM® s.r.l.

Via Scozia, 64026 Roseto degli Abruzzi (TE) Italy
Tel. +39 0858930745 Fax +39 0858930330 www.liofilchem.com

